

Application Note **31309**

Mini-disk 525

Extraction of EPA Method 525 Semi-Volatile Organic Compounds in Water using Mini-Disks and Presto Automated SPE Extractor by Maryland Department of Health (MDH)

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Keywords

EPA Method 525, semi-volatile, Minidisk, Presto Automated SPE, water



ABSTRACT

Solid phase extraction (SPE) is commonly used in the analysis of semi-volatile organic compounds (SVOCs) in drinking water. To capture the full advantages of SPE disks and cartridges, PromoChrom developed a suite of Mini-disks which come in a format similar to a 30 mm syringe filter and have a cross-sectional area 5 times that of a 6 mL SPE cartridge. The increased cross-sectional area and optimized sorbent properties enable the Mini-disks to work with much higher flow rates than SPE cartridges while consuming less solvent than SPE disks. Its compact size makes it easily adaptable to any PromoChrom automated extraction system, especially when paired with the latest Presto automated SPE system, which significantly shortens

the time needed for sample loading. This application note demonstrates the extraction of 22 SVOCs in water using our MD-525-30 Mini-disk with the Presto automated SPE system validated by MDH.

INTRODUCTION

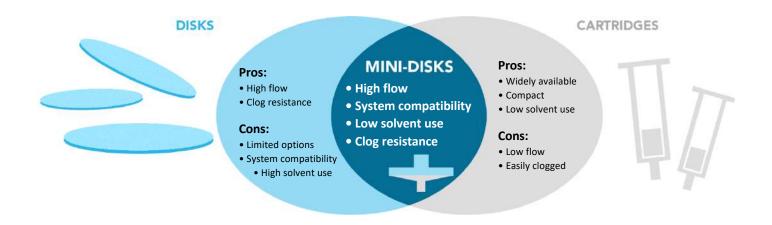
Pesticides are widely used in agriculture to protect crops and improve yields, but their persistence in the environment can pose risks to water quality and public health. Detecting and quantifying trace levels of pesticides in water is essential for ensuring compliance with environmental regulations and safeguarding drinking water supplies.

As a major regional monitoring and testing government body, 'Maryland Department of Health-Laboratories Administration (MDH)' performs a wide range of sample testing, which includes testing for trace organic pesticides in tap water, well water and drinking water samples per EPA Method 525.2 to ensure the well-being of citizens. They are constantly looking for innovative techniques to optimize their workload and improve their performance.



EPA method 525.2 is a method for the determination of organic compounds in drinking water by extracting with liquid solid extraction (LSE) and identifying/quantifying by Gas Chromatography-Mass Spectrometry (GCMS). Per EPA Method 525, 1 L samples are extracted with LSE cartridges, SPE disks, or SPE cartridges. Even though SPE disks offer faster sample loading speed and better tolerance to clogging, they require more solvent usage and have limited market options and compatibility with automated systems. SPE cartridges are more affordable, widely available and compatible with most extraction platforms.

In this application note, we bring forth a novel Mini-disk that combines the advantages and addresses the shortcomings of cartridges and disks. The diameter of the Mini-disk measures only 30 mm and is packaged to fit standard luer slip connections, making it suitable for cartridge-based automated SPE systems and vacuum manifolds. In combination with the PromoChrom Presto, a 50 mL/min flow rate can be achieved for the conditioning step, with a 35 mL/min sample loading rate and 5 mL/min for the final elution. The total extraction time is within 90 minutes and the total collection solvent volume is 19 mL (less than 50 mL after drying).



This application note demonstrates a series of tests that MDH has performed using MD-525-30 Mini-disk with the Presto automated solid-phase extraction system to extract 22 SVOCs from water samples for analysis.

MATERIALS

- Standards for sample spiking and GCMS analysis: Alachlor, Aldrin, Atrazine, Benzo(a)pyrene, Bis(2-ethylhexyl) Adipate, Bis(2ethylhexyl) Phthalate, Butachlor, Cis-Chlordane, Dieldrin, Endrin, Heptachlor, Heptachlor epoxide, Hexachlorobenzene, Hexachlorocyclopentadiene, Lindane, Methoxychlor, Metolachlor, Metribuzin, Propachlor, Simazine, Trans-Chlordane, Trans-Nonachlor (see Appendix Table 1 for details)
- Internal standards: Acenaphthalene-d10, Phenanthrene-d10, Chrysene-d12 (see Appendix Table 1 for details)
- System Monitoring Compound (Surrogate): Perylene-d12 (see Appendix Table 1 for details)
- Mini-disk: PromoChrom Mini-disk packed with mixed-mode polymers and capable of extracting both hydrophilic and hydrophobic organic compounds in water (Cat. No.: MD-525-30).
- Equipment for Extraction: PromoChrom Presto automated SPE system.
- Instrument for analysis: Agilent GCMS system including a 7890B GC with split/splitless inlet, a 7693 autosampler and a 5977A Inert MSD. The column is an Agilent J&W HP-5MS 30m x 0.25mm x 0.25 μ m.



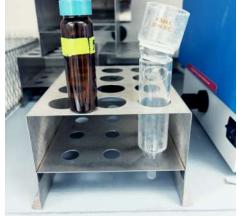




Figure 1: Presto SPE

Figure 2: Collection Drying Apparatus

Figure 3: MD-525-30 Mini-Disk

METHOD

Sample Extraction

Sample extraction and GCMS analysis were performed by MDH.

Prior to the extraction, eight samples consisting of 950 mL of reagent water in 1 liter amber glass bottle were prepared by addition of 40-50 mg of sodium sulfite as preservative, and then acidified to pH <2 using 6N hydrochloric acid. 5 mL of methanol was then added to the sample to assist dispersing of the standards. An aliquot of 40 μ L stock standard solution (containing 2 μ g for each analyte, added to Laboratory Fortified Blank (LFB), Laboratory Fortified Sample Matrix (LFM) samples), 100 μ L stock Internal Standard (IS) and Surrogate Standard (SS) (containing 5 μ g for each IS/SS) were added to the water samples.

Samples were extracted using the method shown below:

Procedures programmed on the Presto automated SPE system

Solvent 1 = Methanol (MeOH), Solvent 2 = Water, Solvent 3 = Ethyl Acetate (EtOAc), Solvent 4 = Methylene chloride (DCM);

W1 = Aqueous waste, W2 = Organic waste

Action	Inlet 1	Flow	Volume	Description	
Elute W2	Solvent 4	50 mL/min	10 mL	Condition Mini-disks with 10 mL of DCM	
Elute W2	Solvent 3	50 mL/min	10 mL	Condition Mini-disks with 10 mL of EtOAc	
Wait	Time based		1 min	Allow 1 minute soak	
Elute W2	Solvent 3	50 mL/min	5 mL	Condition Mini-disks with 5 mL of EtOAc	
Air-Purge W2	Air	50 mL/min	10 mL	Purge residual solvent from Mini-disks to waste 2	
Elute W2	Solvent 1	50 mL/min	10 mL	Condition Mini-disks with 10 mL MeOH	
Wait	Time base	d	1 min	Allow 1 minute soak	
Elute W2	Solvent 1	50 mL/min	5 mL	Condition Mini-disks with 5 mL MeOH	
Elute W1	Solvent 2	50 mL/min	10 mL	Condition Mini-disks with 10 mL water	
Add Sample W1	Sample	35 mL/min	10 mL	Load samples at 35 mL/min, to prime the pump	
Add Sample+ W1	Sample	35 mL/min	1060 mL	Load samples at 35 mL/min through presto pump, using 1060 mL to ensure all sample liquid in the bottles are loaded	
Elute W1	Solvent 2	50 mL/min	20 mL	Wash Mini-disks with 20 mL water	
Air-Purge W1	Air 70 mL/min		10 mL	Remove water droplets from the Mini-disks	
Blow N2	Time based		20 min	Use nitrogen to dry the Mini-disks @ 30psi which gives about ~4-4.5 L/min flow rate for 8 channels	
Rinse	Solvent 3	70 mL/min	5 mL	Rinse sample bottle with 5 mL EtOAc	
Air-Purge R	Air	70 mL/min	5 mL	Purge rinse lines	
Collect 2	Sample	5 mL/min	5 mL	Collect rinsate into Fraction 2	
Wait	Time base	d	1 min	Allow liquid to settle for 1 minute	
Rinse	Solvent 4	70 mL/min	5 mL	Rinse sample bottle with 5 mL DCM	
Air-Purge R	Air	70 mL/min	5 mL	Purge solvent out of rinse line	
Collect 2	Sample	5 mL/min	5 mL	Collect rinsate into Fraction 2	
Wait	Time based		1 min	Allow liquid to settle for 1 minute	
Rinse	Solvent 4	Solvent 4 70 mL/min		Rinse the sample bottle with 3 mL DCM	
Air-Purge R	Air	70 mL/min	5 mL	Purge rinse lines	
Collect 2	Sample	5 mL/min	10 mL	Collect rinsate into fraction 2	
Collect 2	Solvent 4	5 mL/min	3 mL	Elute the Mini-disks directly with DCM and collect into fraction 2	

Action	Inlet 1	Flow	Volume	Description
Wait	Time base	ed	1 min	Allow liquid to settle for 1 minute
Collect 2	Solvent 3	5 mL/min	3 mL	Elute the Mini-disks directly with EtOAc and collect into fraction 2
Air-Purge 2	Air	50 mL/min	15 mL	Push any remaining sample into fraction 2

The extraction took 81 minutes for a batch of 8 samples. The final extract in the collection vial was about 19 mL. 6-10 g of sodium sulfate was then added to the collection vial and the vial was vortexed. The mixture was then passed through 3-5 g of sodium sulfate and washed with 3 \times 10 mL of DCM. The sample was concentrated to <1 mL (above 0.5 mL mark) using an evaporator with an endpoint detection sensor. Then, 10 μ L of Recovery standard (RS) was added, and the volume was adjusted to 1.0 mL prior to GC-MS analysis.

Evaporator parameters: 45° C with a N_2 pressure setting of 9-10 psi. The N_2 pressure setting will start low at about 3-5 psi to avoid splashing when the liquid level is high in the collection vials.

GCMS Analysis

The parameters for GCMS analysis are listed below:

Parameters for GCMS analysis

Injection Volume	2 μL
Inlet	270 °C; Pulsed splitless injection mode
Oven Temperature Program	80 °C (hold for 0.5 minutes), 18 °C/min to 260 °C 6 °C/min to 310 °C (hold for 1 minute) Total run time: 19.83 mins
Carrier Gas and Flow Rate	Helium at 1.22 mL/min, constant flow
Transfer Line Temperature	280 °C
Ion Source Temperature	230 °C
Quadrupole Temperature	150 °C
Scan	50 to 550 <i>m/z</i>

RESULTS AND DISCUSSION

The recoveries of 22 compounds are summarized in Appendix Table 1. All analytes were spiked at 2 ppb and all recoveries fall within the 70-130% range, which meets the criteria of EPA 525.2 (as shown in Figure 4). The %RSD for most of the compound's are within 10% for 8 replicates conducted over 2 days.

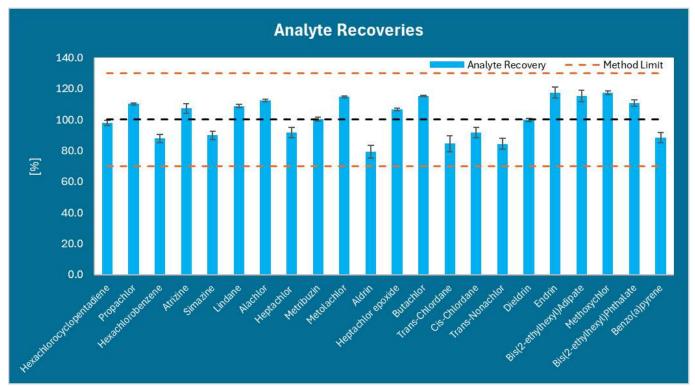


Figure 4: Analyte Recoveries on the Presto-SPE System: The method limits are illustrated as the red dashed line, with %RSD shown as an error bar. The black dashed line is 100%. Note: *Technical Chlordane: average of Trans-Chlordane, Cis-Chlordane and Trans-Nonachlor

The method detection limit (MDL) and reporting limit (RL) are shown in Figure 5. MDL verification was performed by running seven spikes at the following concentrations: $0.5~\mu g/L$ for pesticides and low level (LL) pesticides, and $1.0~\mu g/L$ for high level (HL) pesticides. The lab reagent blank (LRB) are reported to be lower than the detection limits. RL of lindane, Heptachlor, Heptachlor epoxide and Benzo(a)pyrene are $0.1~\mu g/L$ as validated on a separate RL study by 7 replicates spiking at $0.1~\mu g/L$.

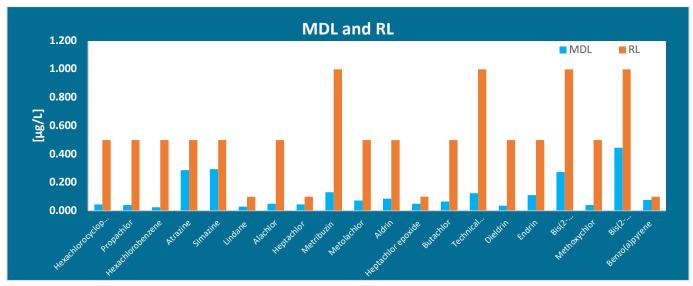


Figure 5: MDL and RL of Analytes on the Pesto-SPE System.

The average recovery of internal standards (IS), surrogate standards (SS) and recovery standards (RS) are in Figure 6. All recoveries also fall within the 70-130% for the EPA requirement. Please note that for EPA 525.2, IS/SS were spiked with 5 μ g per liter of sample (pre-spiking), and RS was added at 5 μ g per mL of the extracted sample in the final micro-vial (post-spiking). The %RSD for most of the compound's are within 10%.

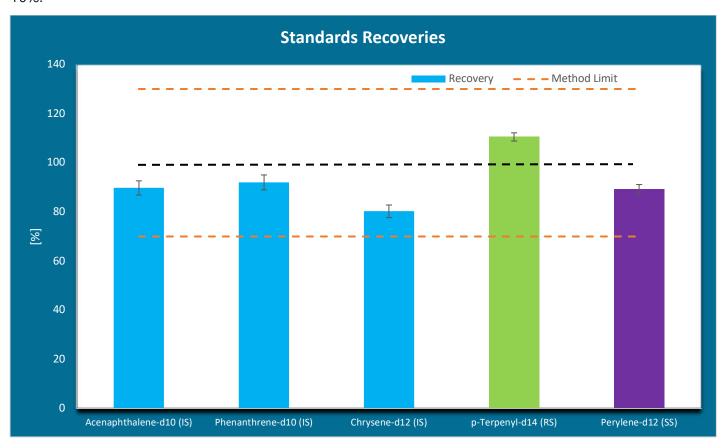


Figure 6: Recoveries of Internal standards (IS), Recovery standard (RS) and Surrogate standard (SS) on the Presto-SPE System with %RSD shown as an error bar. The method limits are illustrated as red dashed line, with %RSD shown as an error bar. The black dashed line is 100%.

CONCLUSION

The PromoChrom MD-525-30 mini-disk in combination with the PromoChrom Presto can effectively extract the 22 SVOCs monitored by MDH while meeting the EPA 525.2 certification criteria. This solution offers an efficient extraction approach taking 81 minutes for 950 mL samples and using a relatively small amount of final collection solvent of 19 mL. MDH diligently serves its community by performing a wide range of tests using automated SPE and developing optimal workflows through automation.

ORDERING INFORMATION FOR MINI-DISKS

Part number	Description	Application
MD-525-30	Packed with 25-um spherical mixed-mode sorbent, pack of 40	For extraction of hydrophobic and hydrophilic compounds.

REFERENCES

 EPA method 525.2, EPA Method 525.2: Determination of Organic Compounds in Drinking Water by Liquid-Solid Extractions and Capillary Colum Gas Chromatography/Mass Spectrometry.

Appendix

Table 1 - Average recoveries of analytes from 8 spiked samples

No.	Analytes	CAS No.	Average	RSD (%)	MDL (μg/L)	RL
1	Hexachlorocyclopentadiene	77-47-4	97.9	2.97	0.044	0.50
2	Propachlor	1918-16-7	110.3	1.19	0.041	0.50
3	Hexachlorobenzene	118-74-1	87.9	5.49	0.024	0.50
4	Atrazine	1912-24-9	107.3	6.00	0.287	0.50
5	Simazine	122-34-9	89.9	5.18	0.294	0.50
6	Lindane	58-89-8	108.8	2.07	0.029	0.10
7	Alachlor	15972-60-8	112.4	1.56	0.049	0.50
8	Heptachlor	76-44-8	91.8	6.42	0.044	0.10
9	Metribuzin	21087-64-9	100.4	2.78	0.130	1.00
10	Metolachlor	51218-45-2	114.8	1.21	0.072	0.50
11	Aldrin	309-00-2	79.2	8.25	0.086	0.50
12	Heptachlor epoxide	1024-57-3	106.5	1.66	0.049	0.10
13	Butachlor	23184-66-9	115.3	1.06	0.065	0.50
14	Trans-Chlordane	5103-74-2	84.5	10.41	0.124*	1.00
15	Cis-Chlordane	5103-71-9	91.7	6.95	0.124*	1.00
16	Trans-Nonachlor	39765-80-5	84.4	7.12	0.124*	1.00
17	Dieldrin	60-57-1	99.9	2.10	0.036	0.50
18	Endrin	72-20-8	117.6	7.03	0.111	0.50
19	Bis(2-ethylhexyl)Adipate	103-23-1	115.3	7.46	0.274	1.00
20	Methoxychlor	72-43-5	117.4	2.53	0.039	0.50
21	Bis(2-ethylhexyl)Phthalate	117-81-7	110.7	3.90	0.446	1.00
22	Benzo(a)pyrene	50-34-8	88.4	6.84	0.076	0.10
Acce	ptance Criteria		70-130	< 30	MDL <rl< td=""><td></td></rl<>	

Note: *Technical Chlordane: average of 3 (Trans-Chlordane, Cis-Chlordane and Trans-Nonachlor)

Table 2 - Recoveries of Internal standards (IS) Recovery standard (RS) and Surrogates (SS)

Analytes	CAS No.	Average Recovery (%)	RSD (%)
Internal st			
Acenaphthalene-d10	15067-26-2	89.8	5.78
Phenanthrene-d10	1517-22-2	92.0	6.08

Analytes	CAS No.	Average Recovery (%)	RSD (%)
Chrysene-d12	1719-03-5	80.3	5.13
Recovery s			
p-Terpenyl-d14	1718-51-0	110.5	3.35
System Monitori			
Perylene-d12	1520-96-3	89.3	3.77
Acceptance Criteria	70-130		

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